

Patient skin fibroblasts: A versatile tool for identification of novel muscular dystrophy disease genes

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The Dystroglycanopathies:
A Patient and Family Conference
08-18-2012





Project 2:
Clinical Studies



Project 1:
**Translational
Lab Research**

Iowa

**Wellstone
Muscular Dystrophy
Research Center**

SEN. PAUL D. WELLSTONE
MD COOPERATIVE
RESEARCH
CENTER



**Muscle Disease
Neuropathology
Conference**

Core A:
Administration



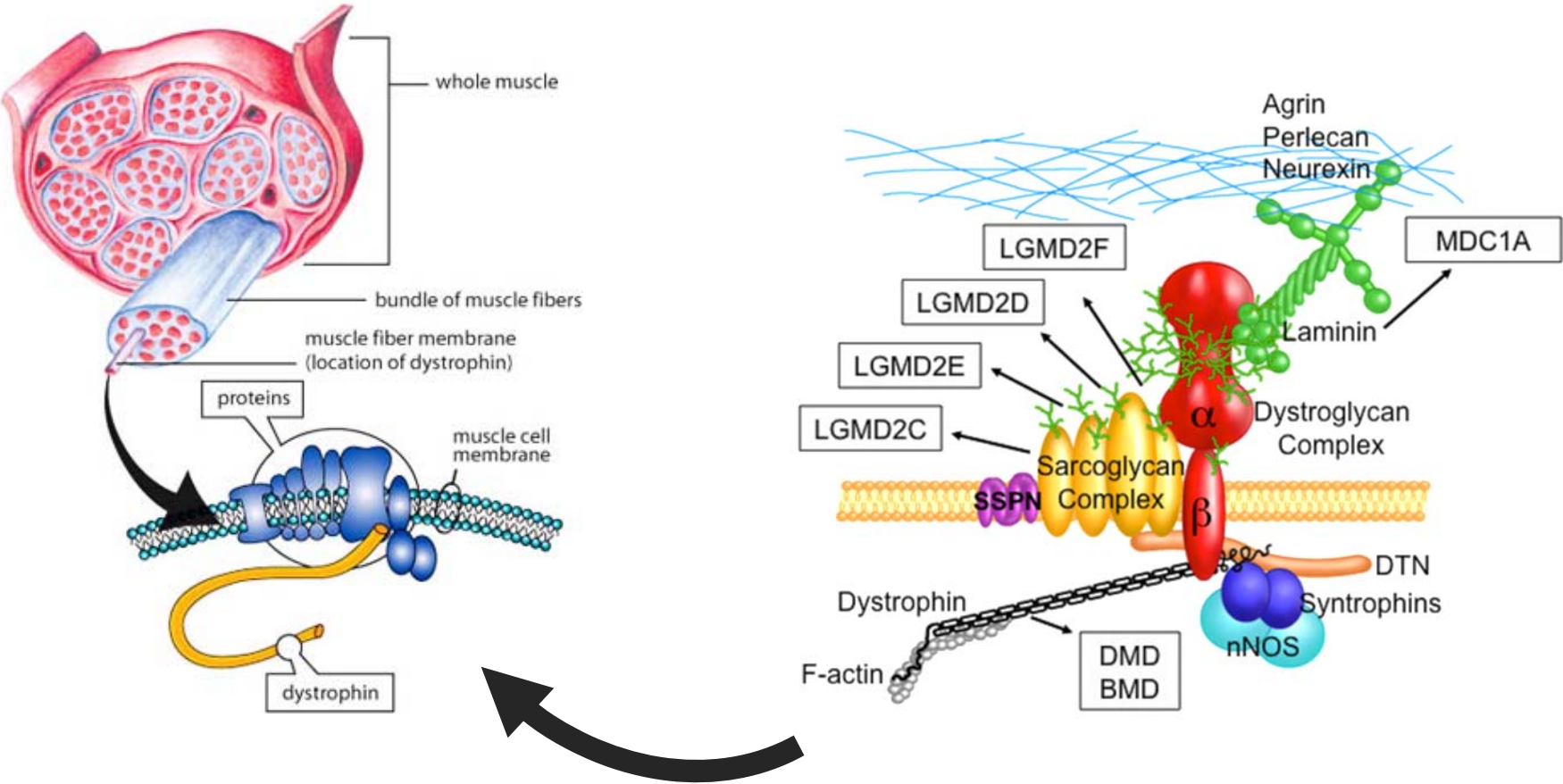
Core B:
**Muscle Tissue and Cell Culture
Repository**



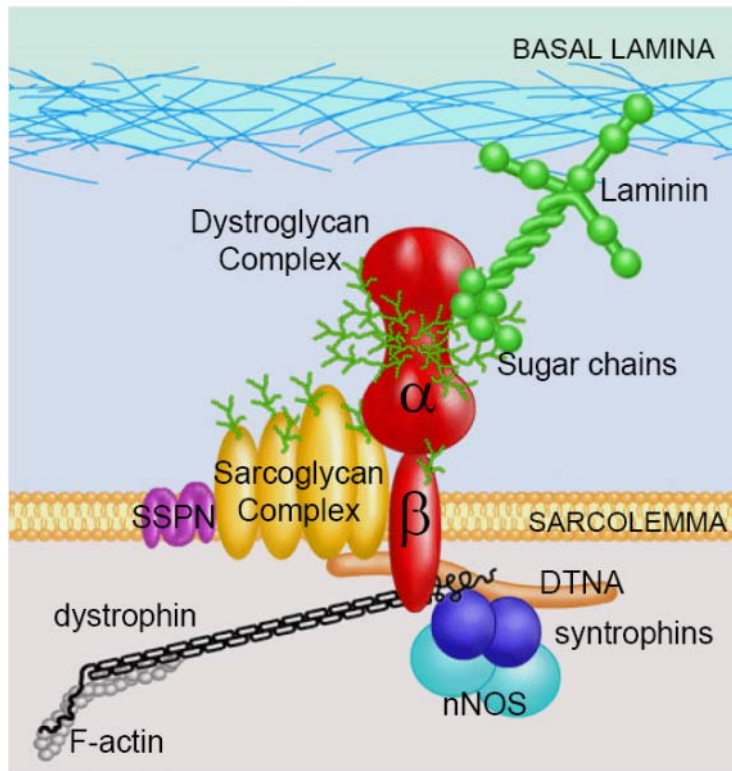
Core C:
Education



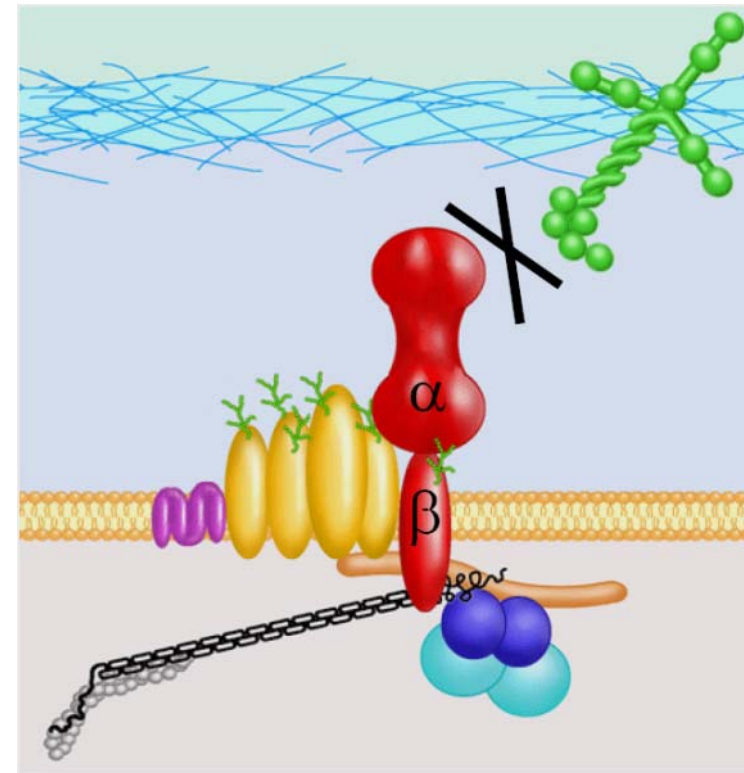
Dystrophin Glycoprotein Complex



Loss of α -Dystroglycan functional glycosylation results in congenital muscular dystrophy

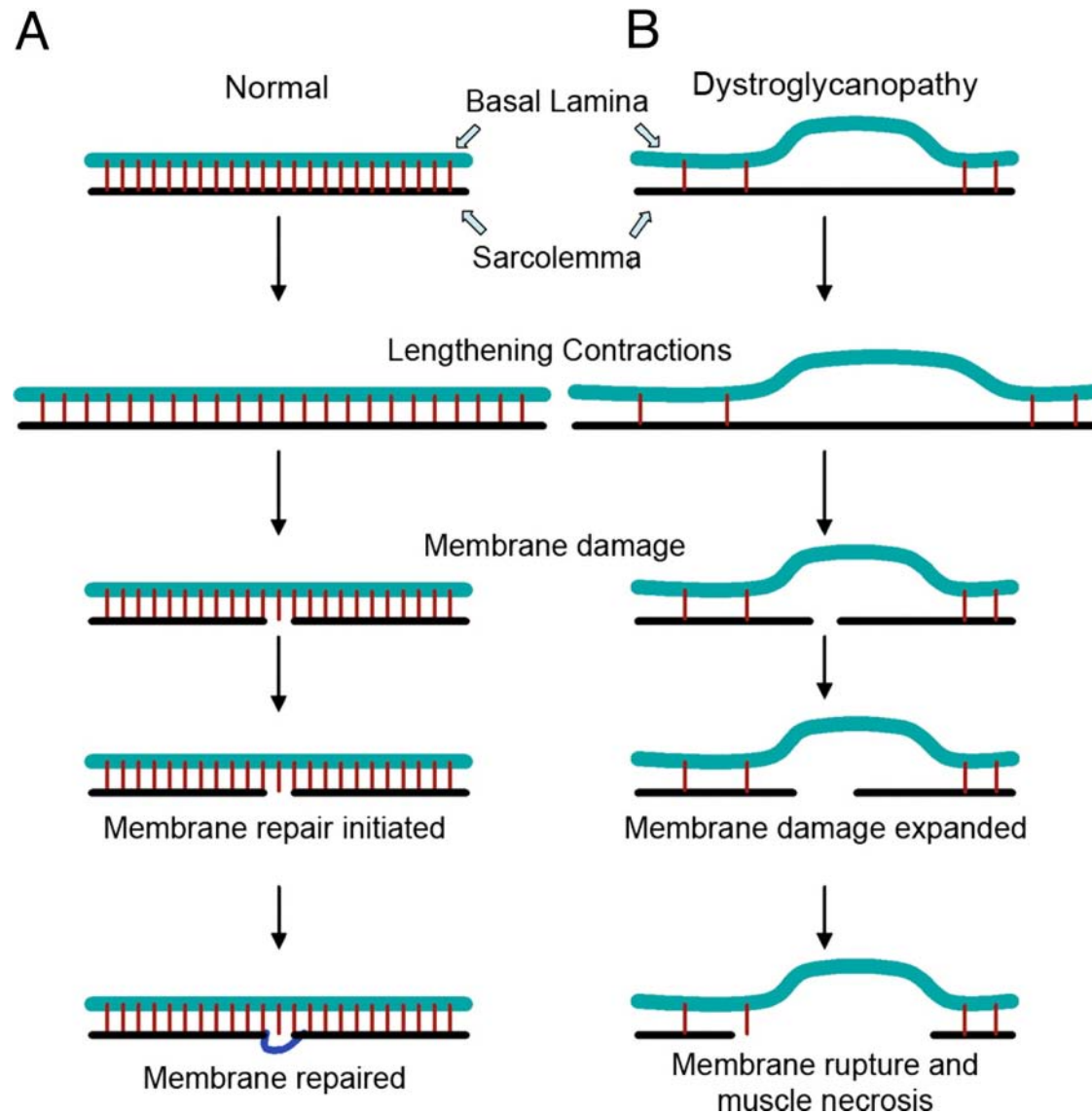


Normal

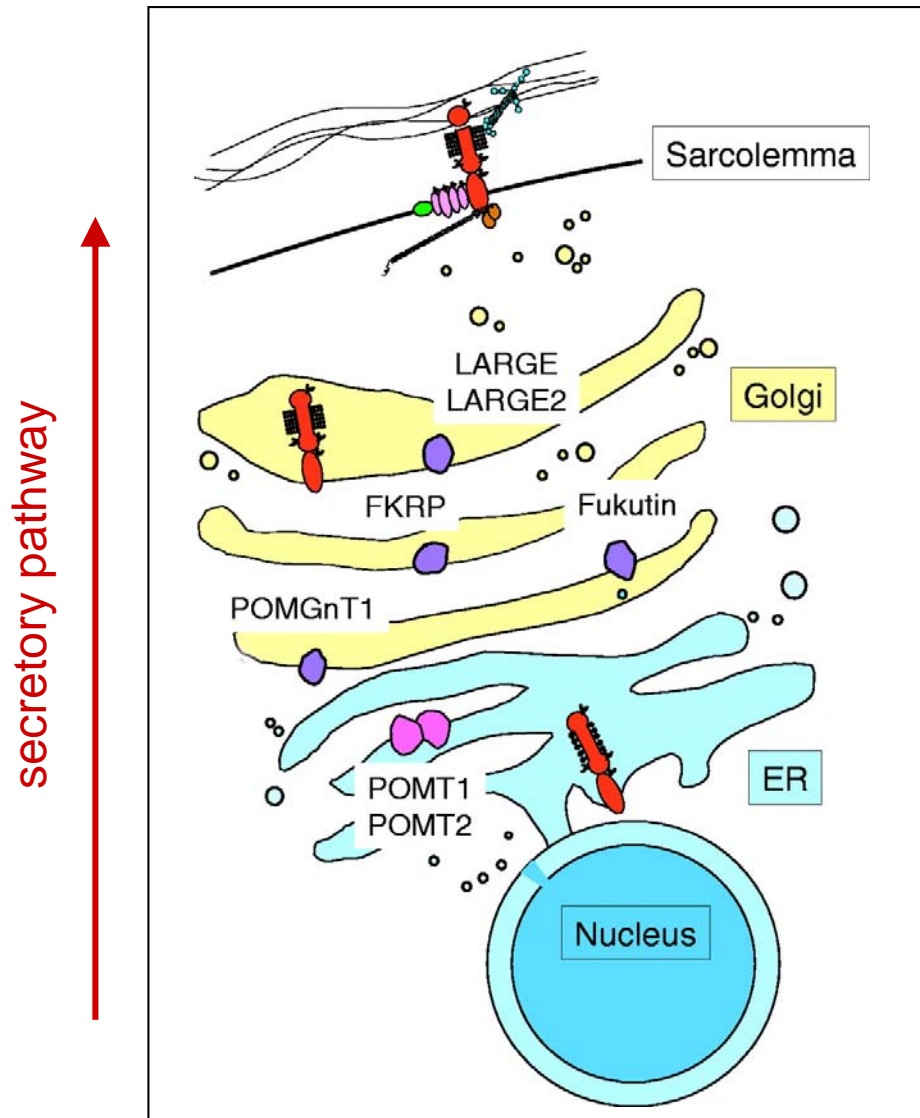


Walker-Warburg syndrome (WWS)
Muscle-eye-brain disease (MEB)
Fukuyama congenital muscular dystrophy (FCMD)
MDC1C/1D
Limb girdle muscular dystrophy (LGMD)2I/2K/2M/2N
Large^{myd} mouse

A proposed mechanism for the basal-lamina-mediated prevention of membrane damage during lengthening contractions.



Glycosylation of α -dystroglycan



6 genes known to be involved in α -dystroglycan glycosylation

Endoplasmic reticulum

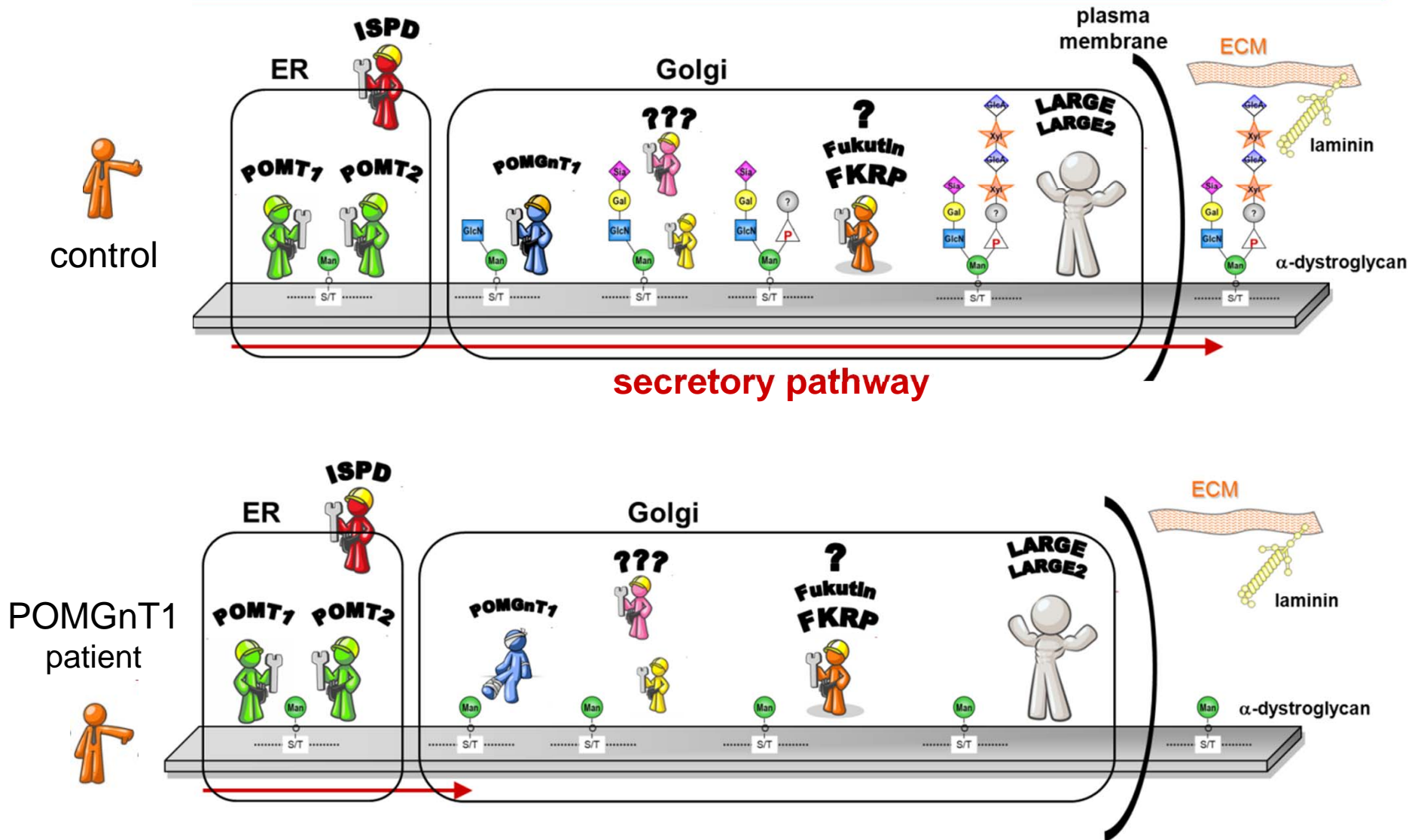
- POMT1
- POMT2

Golgi

- POMGnT1
- FKRP
- Fukutin
- LARGE1

Glycosylation happens during secretion along the secretory pathway

The assembly line - a simple model for α -dystroglycan glycosylation



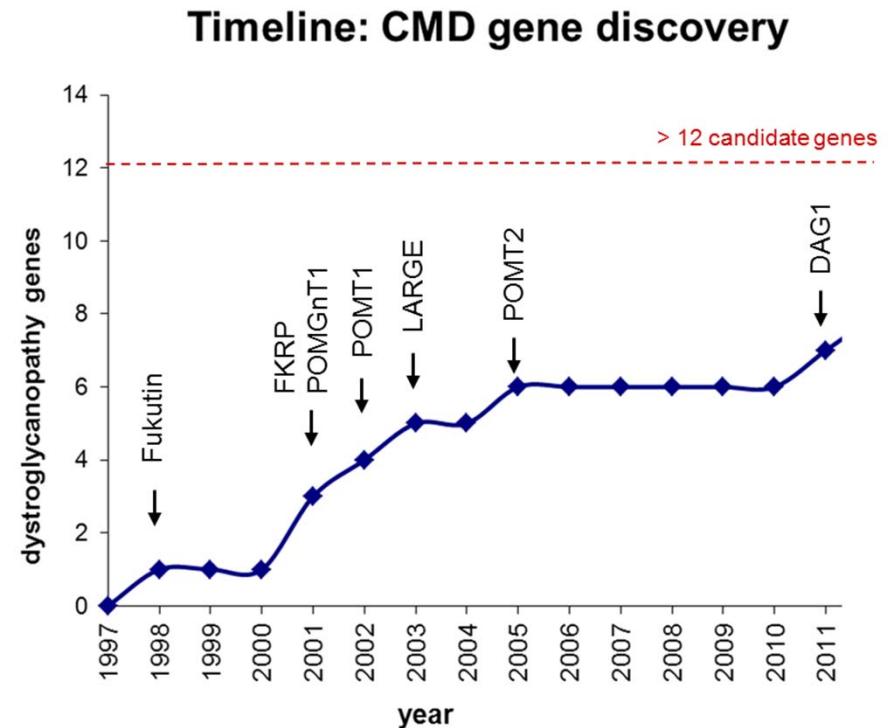
each glycosyltransferase / worker has a very specific skill set to perform only one specific task

Dystroglycanopathy candidate genes

primary dystroglycanopathy: dystroglycan (DAG1) defect, 1 patient identified

secondary dystroglycanopathy: 6 known/putative genes causing

- ***POMT1*** (9q34.1)
- ***POMT2*** (14q24.3)
- ***POMGnT1*** (1p34.1)
- ***FKRP*** (19q13.32)
- ***Fukutin*** (9q31)
- ***LARGE1*** (22q12.3)



Currently only 50% of dystroglycanopathy patients can be genetically diagnosed

Clinical diagnosis



Patient Analysis

Blood

Skin Biopsy

Muscle Biopsy

Lymphocyte

Histology
Immunofluorescence

DNA

Fibroblast

Fibroblast/Myoblast

DNA/RNA

Protein

DNA/RNA

Protein

Sequencing

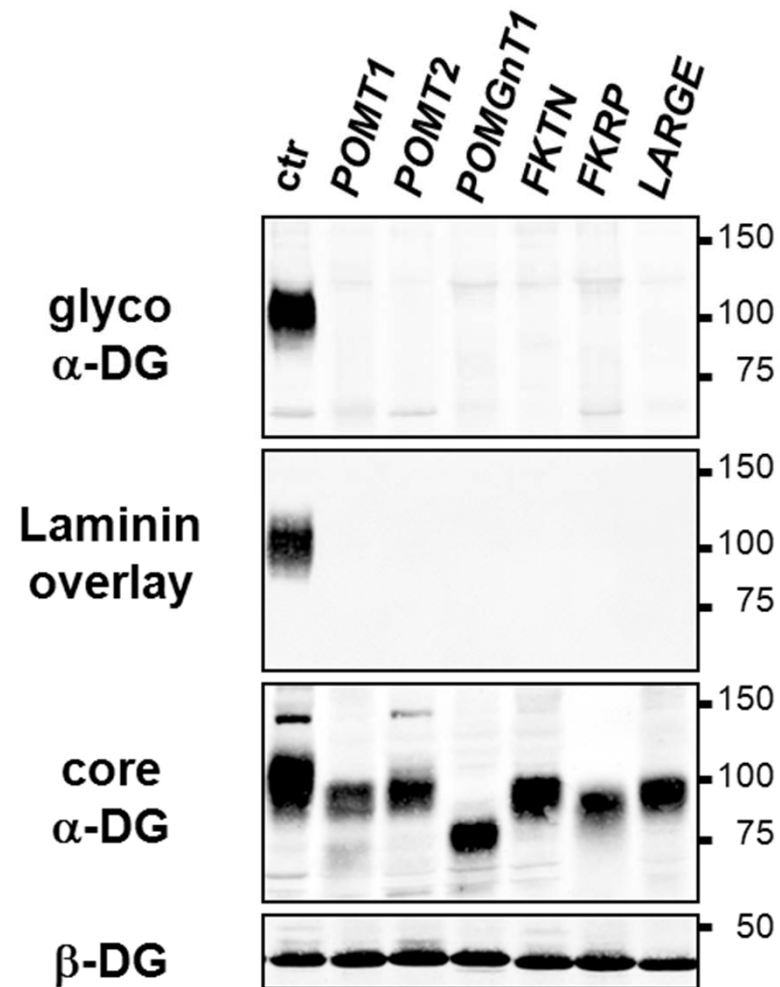
Sequencing/PCR

Western Blot

Sequencing/PCR

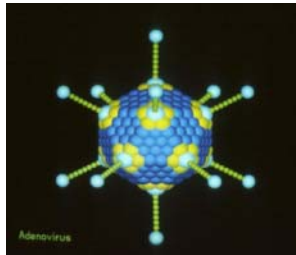
Western Blot

α -DG glycosylation defect in dystroglycanopathy patient skin fibroblasts

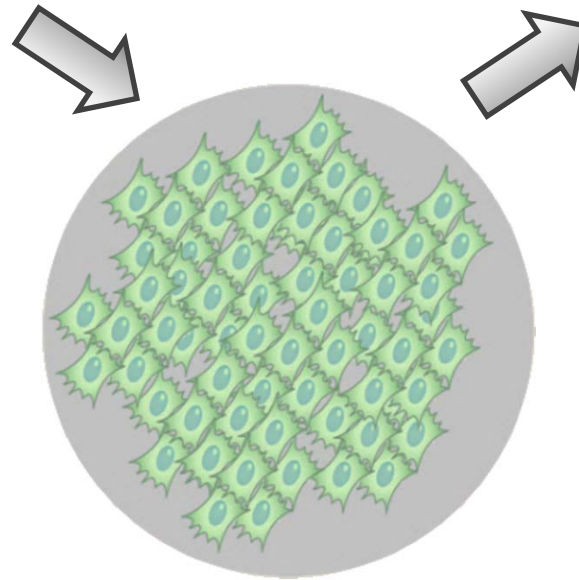


Complementation assay

Adenovirus



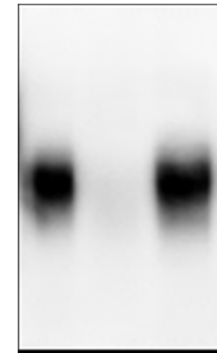
candidate genes



Patient Fibroblasts

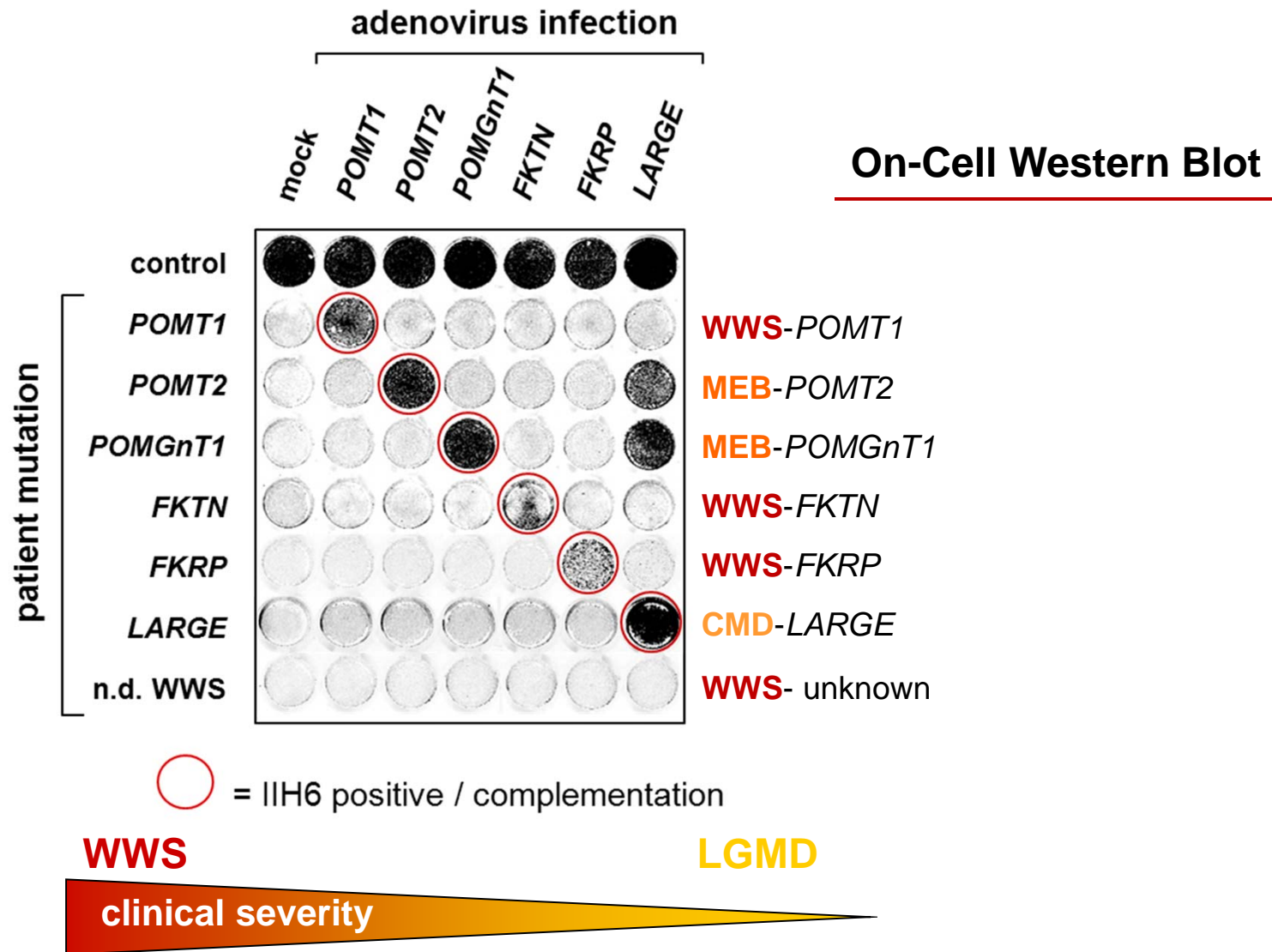
Western Blot

control
WWS
- + complementation

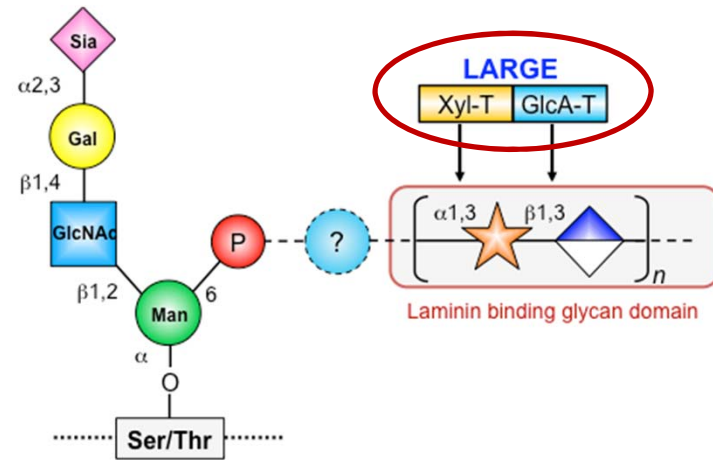
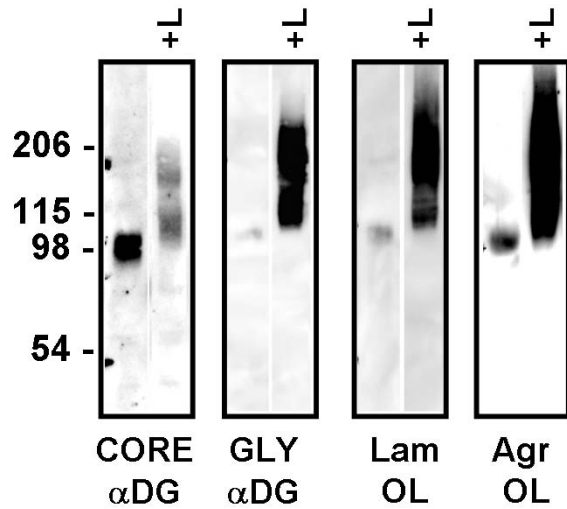


Return of Glycosylation ?

Ability of LARGE to hyperglycosylate α -dystroglycan correlates with the severity of the clinical phenotype

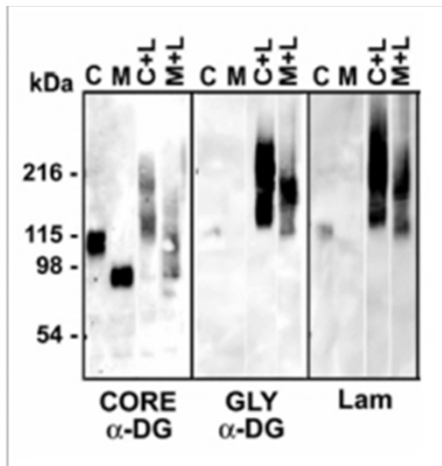


LARGE can bypass α -DG glycosylation defects in CMD patients



The "LARGE" effect

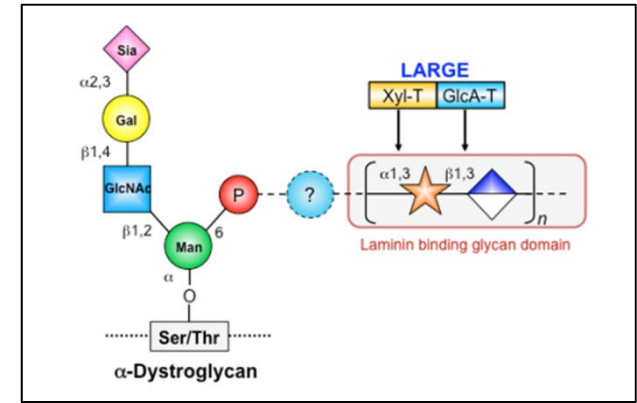
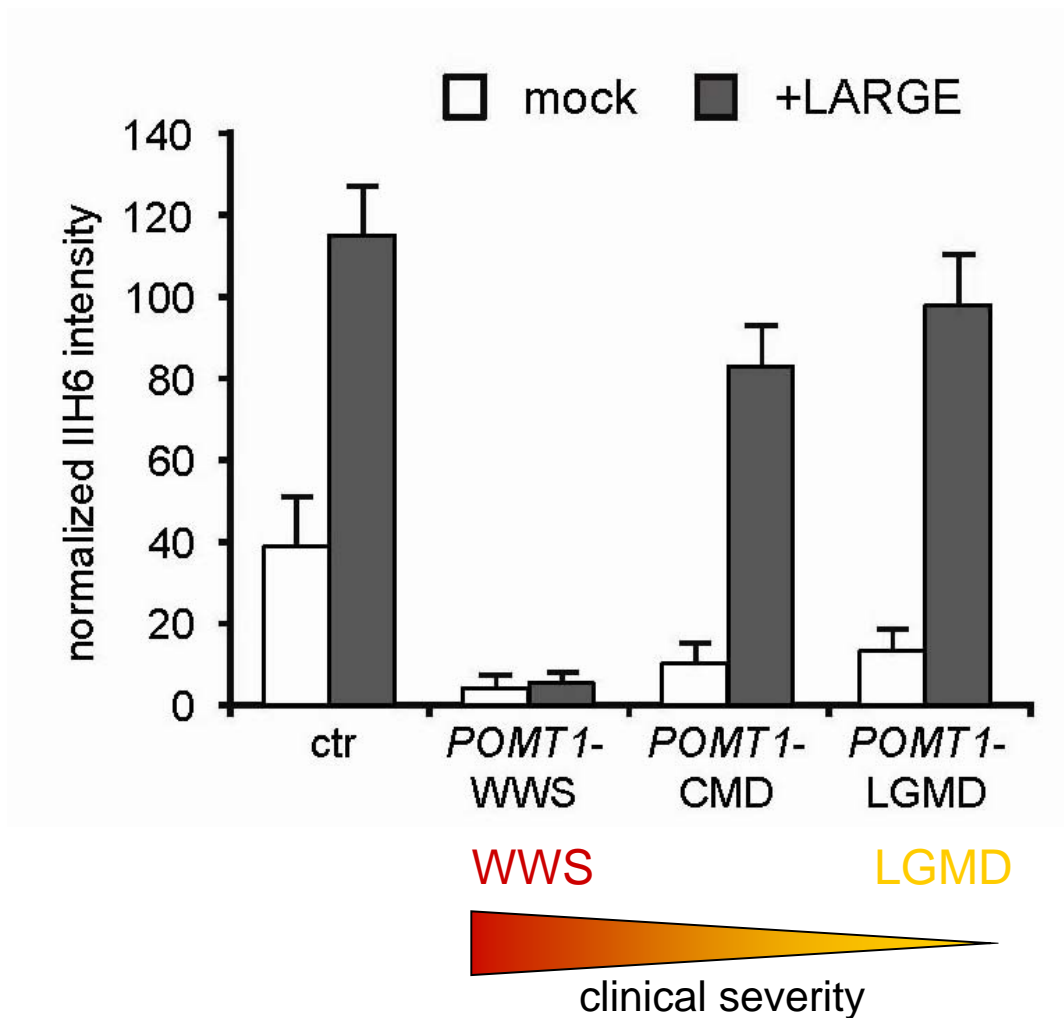
Barresi *et al.*, NatMed (2004)



α -Dystroglycan

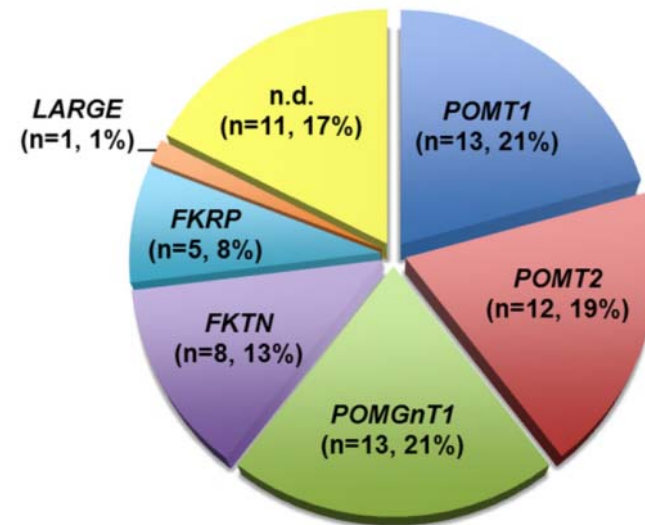
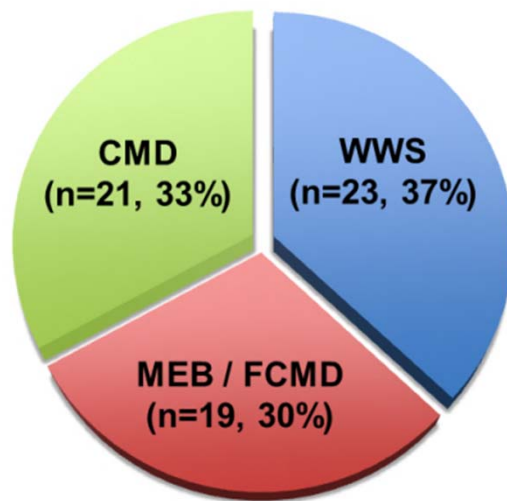
Inamori *et al.*, Science (2012)

Bypass of α -DG glycosylation defects by LARGE correlates with residual activity of impaired CMD gene

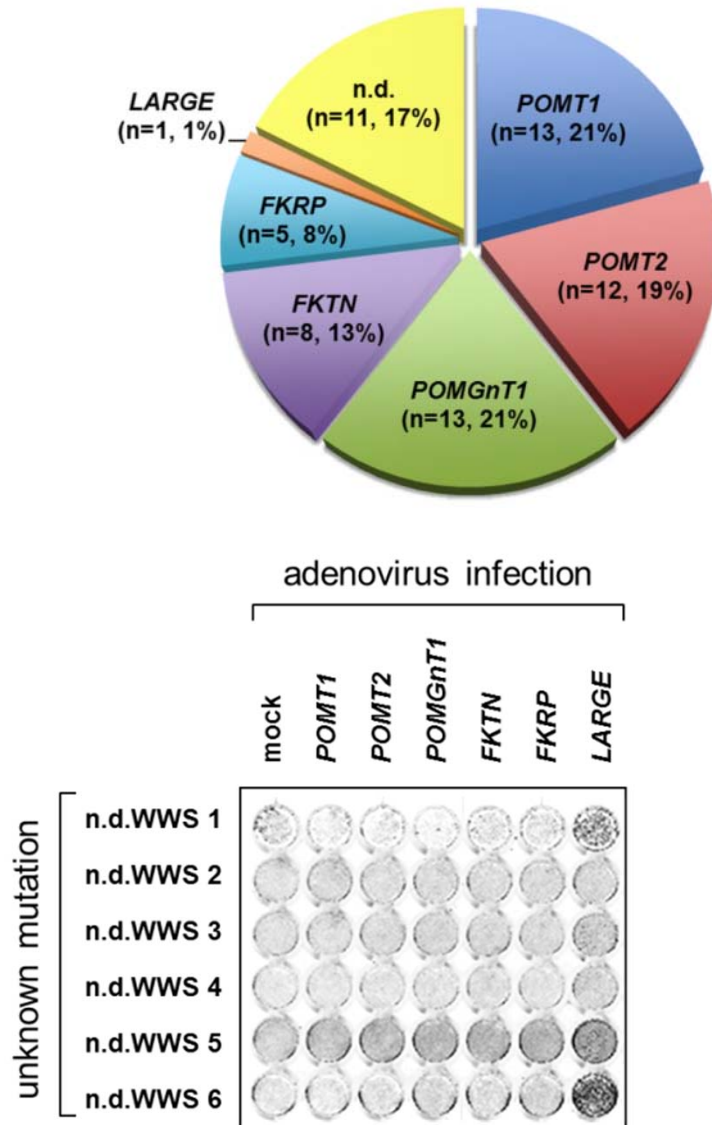


Genetic and Phenotypic distribution of cells analyzed by On-Cell Western Blot complementation (n=63)

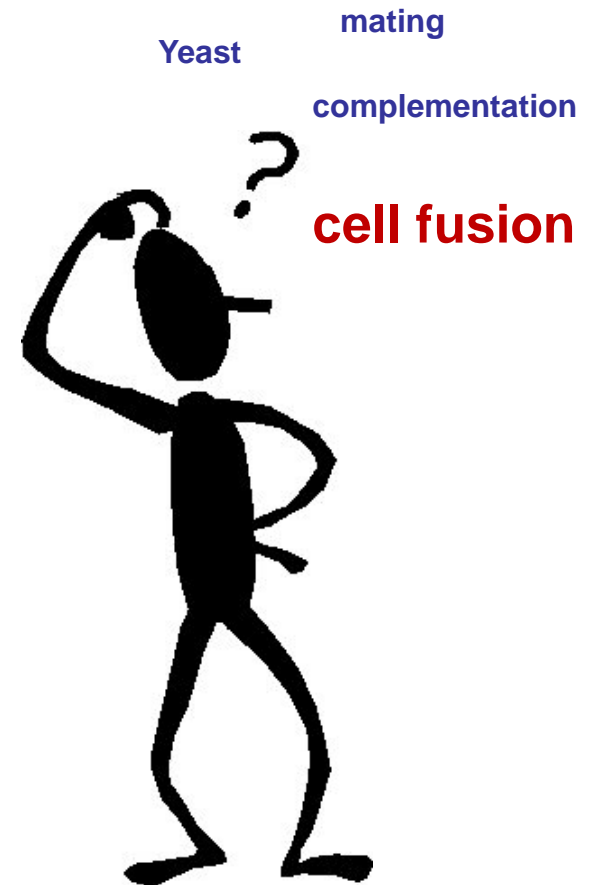
Genetic defect	WWS	MEB/FCMD	CMD	total	total %
<i>POMT1</i>	8		5	13	21%
<i>POMT2</i>	1	6	5	12	19%
<i>POMGnT1</i>	1	9	3	13	21%
<i>FKTN</i>	1	4	3	8	13%
<i>FKRP</i>	1		4	5	8%
<i>LARGE</i>			1	1	1%
n.d.	11			11	17%
total	23	19	21	63	100
total %	37%	30%	33%	100	



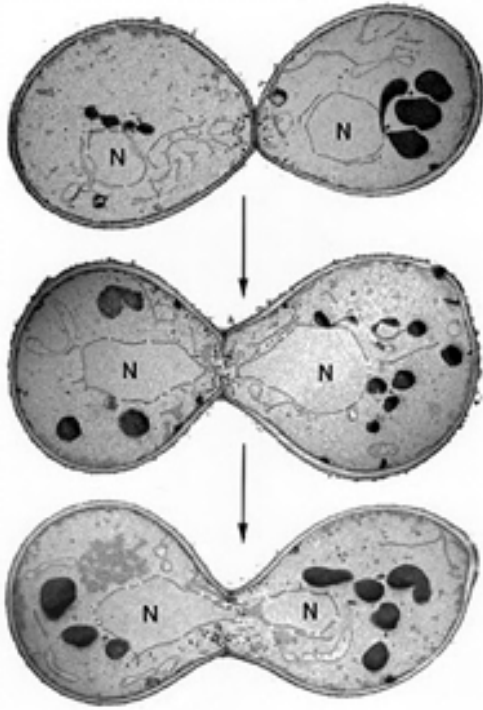
Patient fibroblast complementation assay: unknowns



○ = IIH6 positive / complementation



Yeast cell mating



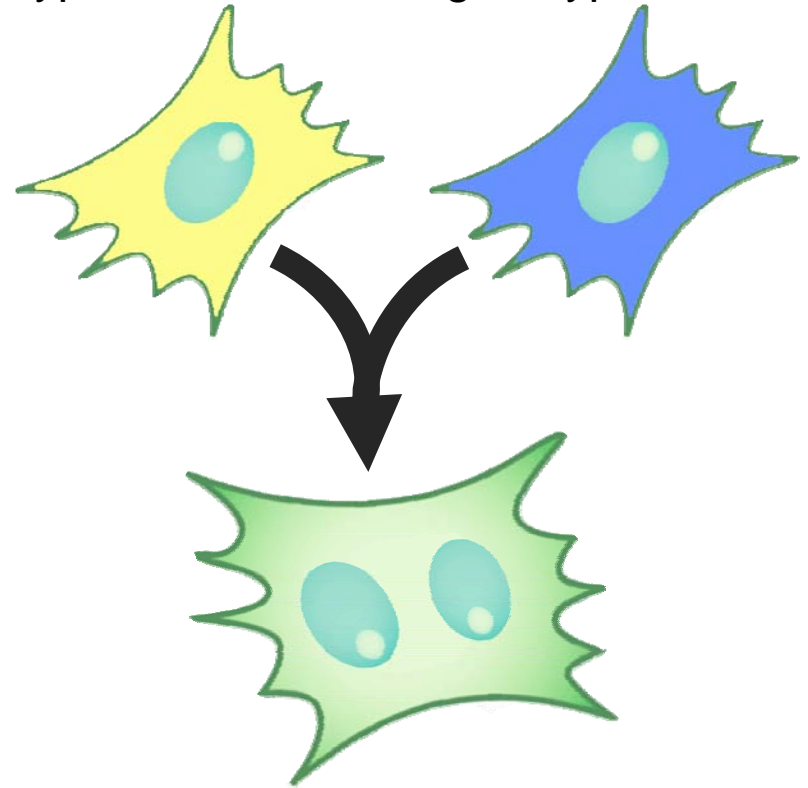
Hypothesis:

Fusion between co-cultured cells harboring recessive mutations with independent genetic defects would result in successful rescue.

PEG induced cell fusion of mammalian cells

IIH6 negative
genotype: aaBB

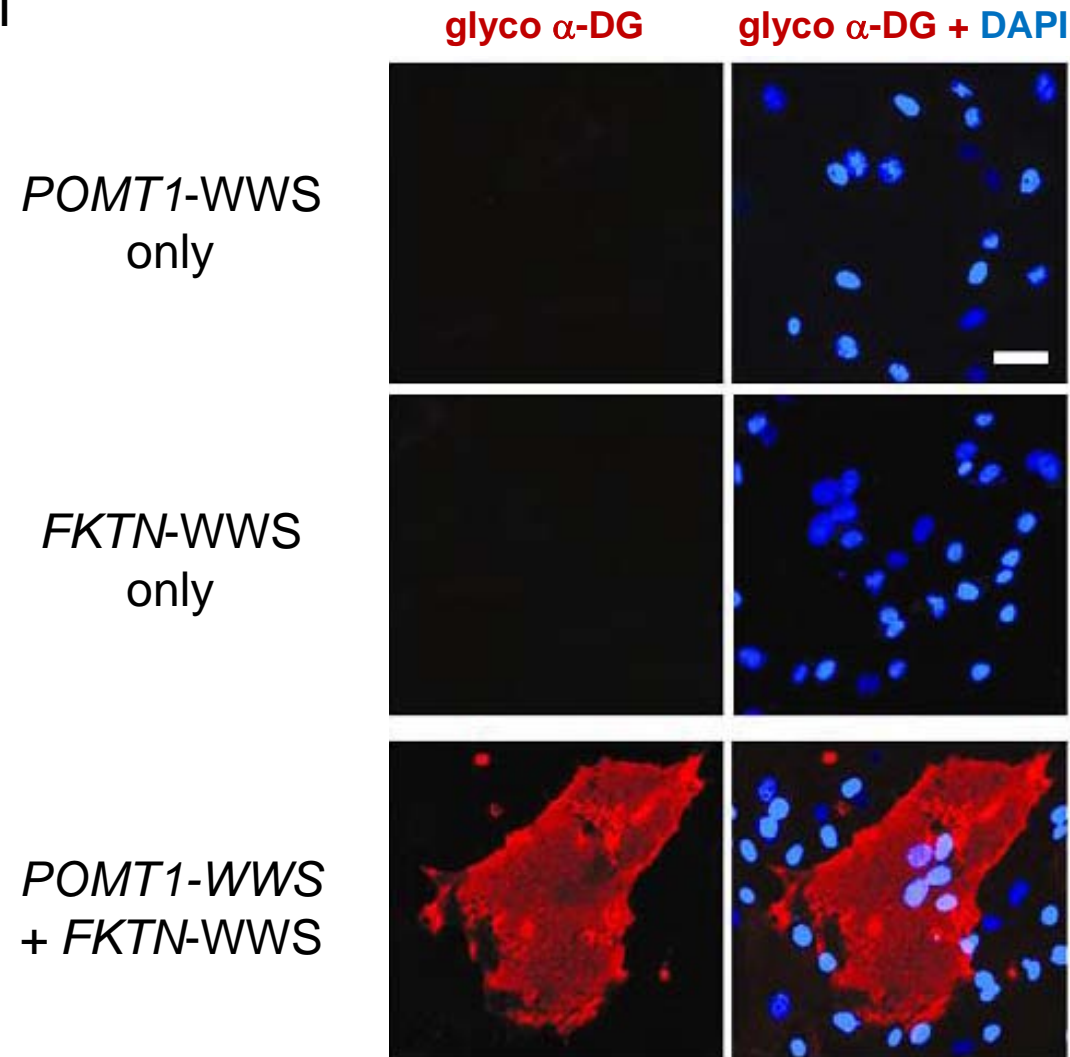
IIH6 negative
genotype: AAbb



IIH6 positive
genotype: aaAABBbb

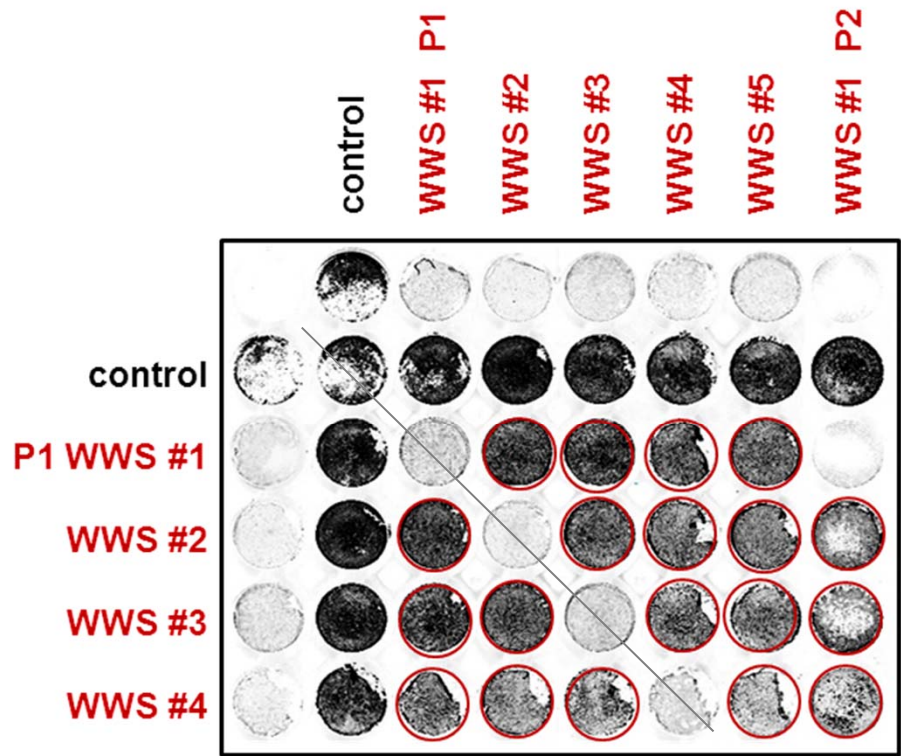
Cell fusion of independent patient fibroblasts restores α -DG glycosylation defect

PEG induced cell fusion assay

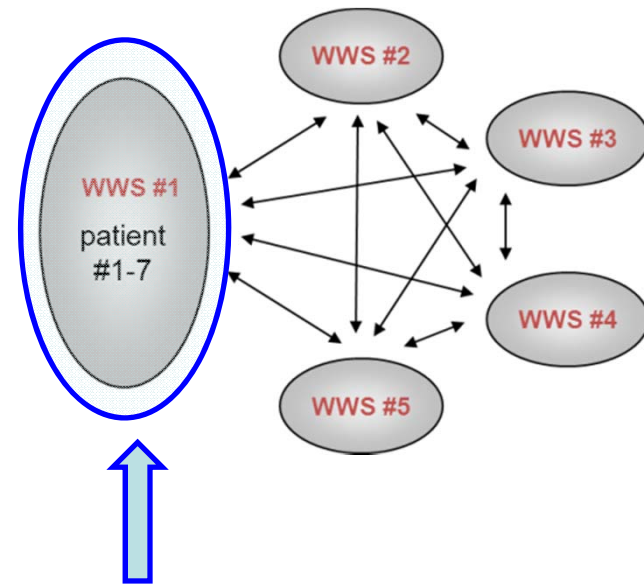


Cell fusion of independent patient fibroblasts restores α -DG glycosylation defect

PEG induced cell fusion assay

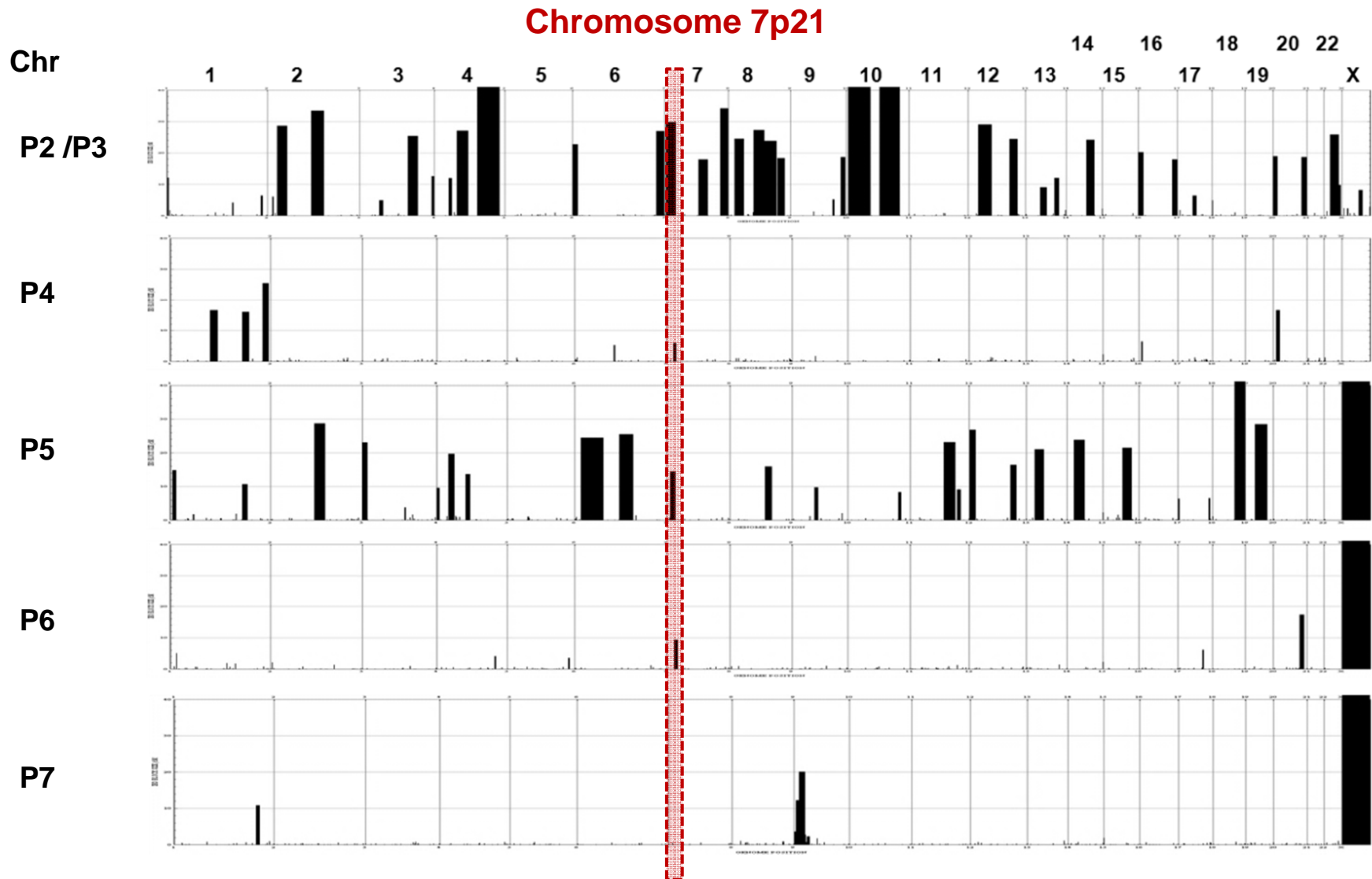


 = IIH6 positive / complementation

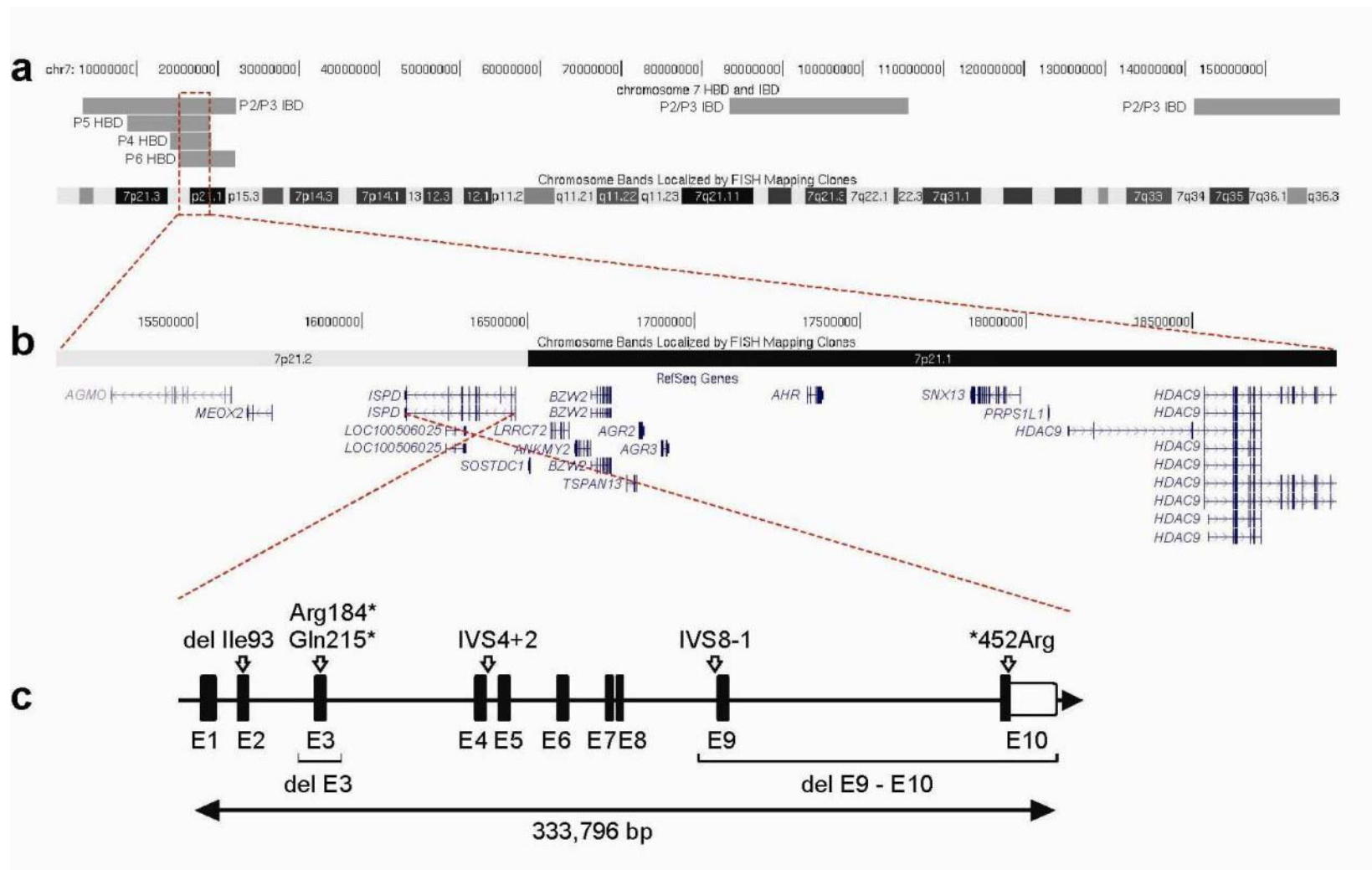


Target group for new gene discovery

Linkage analysis of inbred samples in WWS #1 group

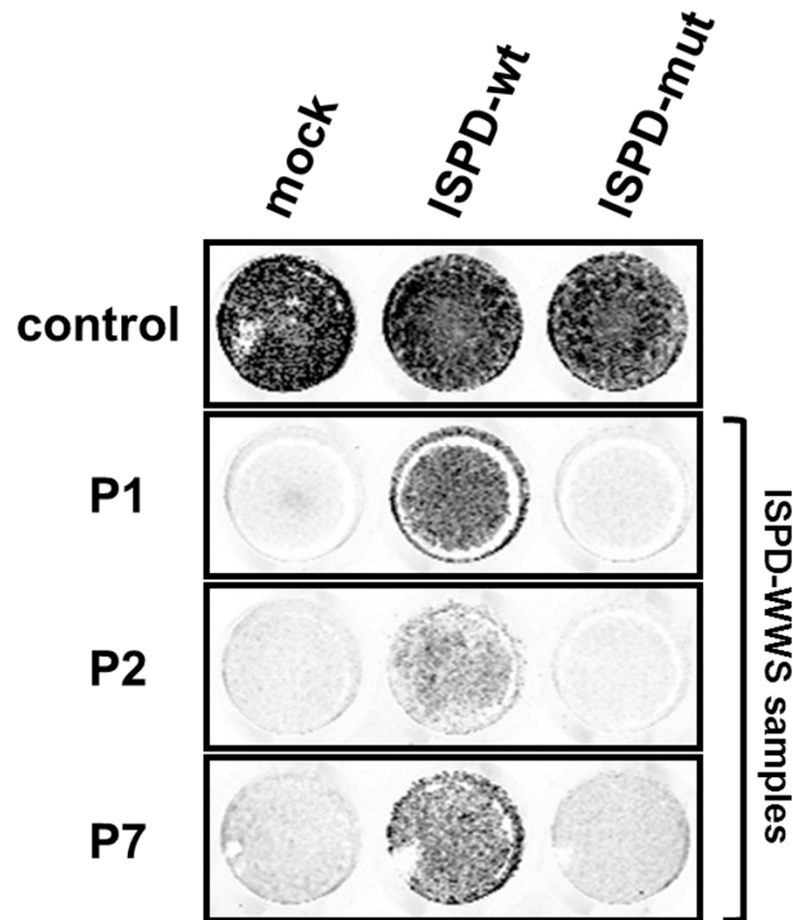


Inbred samples in WWS #1 complementation group have overlapping linkage at Chr. 7p21 and share mutations in *ISPD*



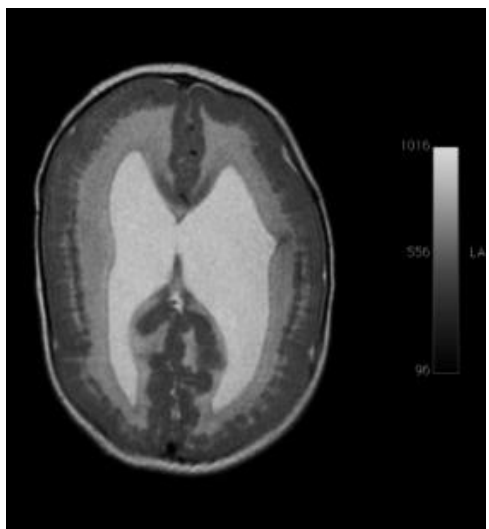
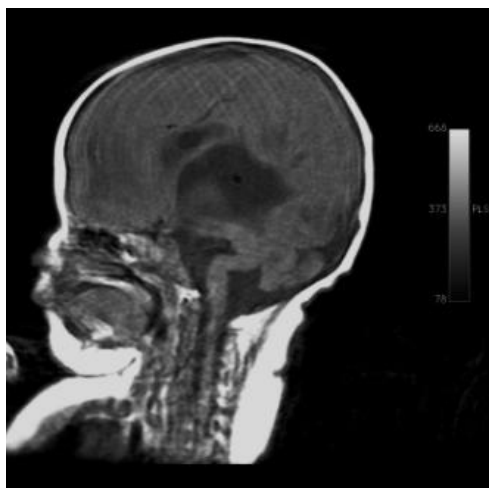
ISPD : Isoprenoid synthase domain containing

Validation of pathogenic *ISPD* mutations with fibroblast complementation

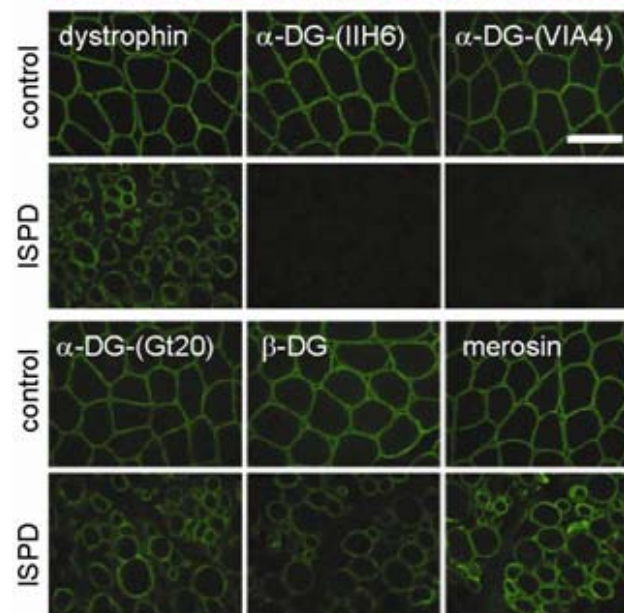
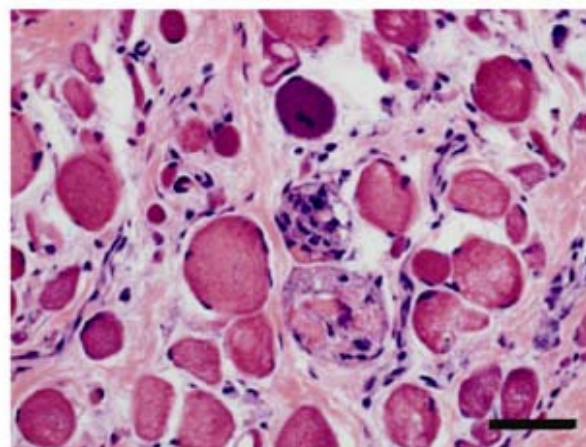


ISPD-WWS patient P1 : clinical presentation

Brain MRI



Muscle biopsy

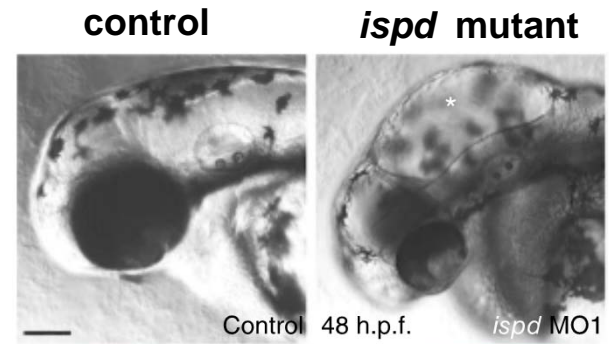


Knockdown of zebrafish *ispd* recapitulates pathological defects of human WWS

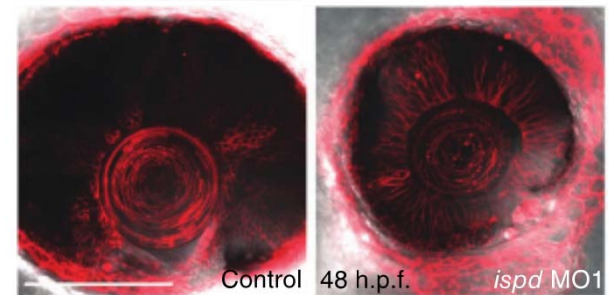
Zebrafish (*Danio rerio*)



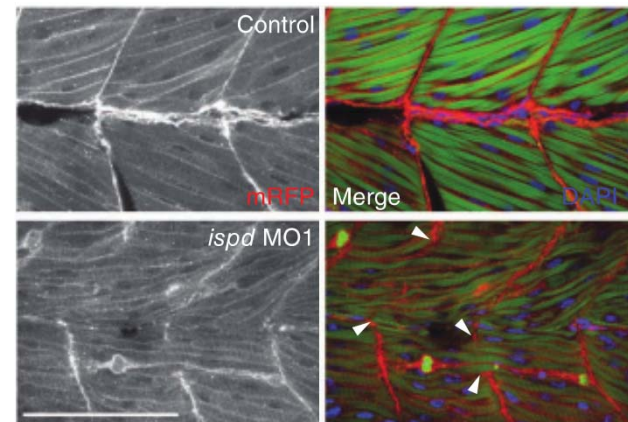
brain



eye

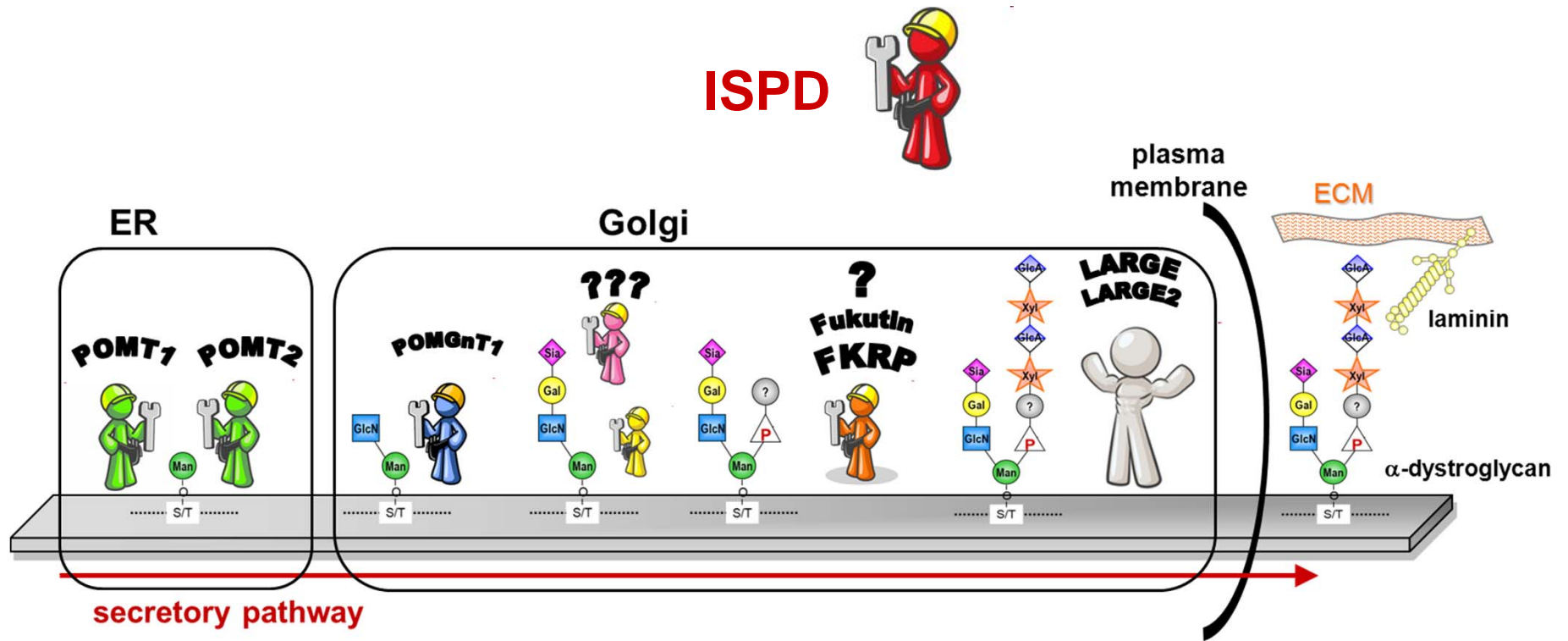


muscle



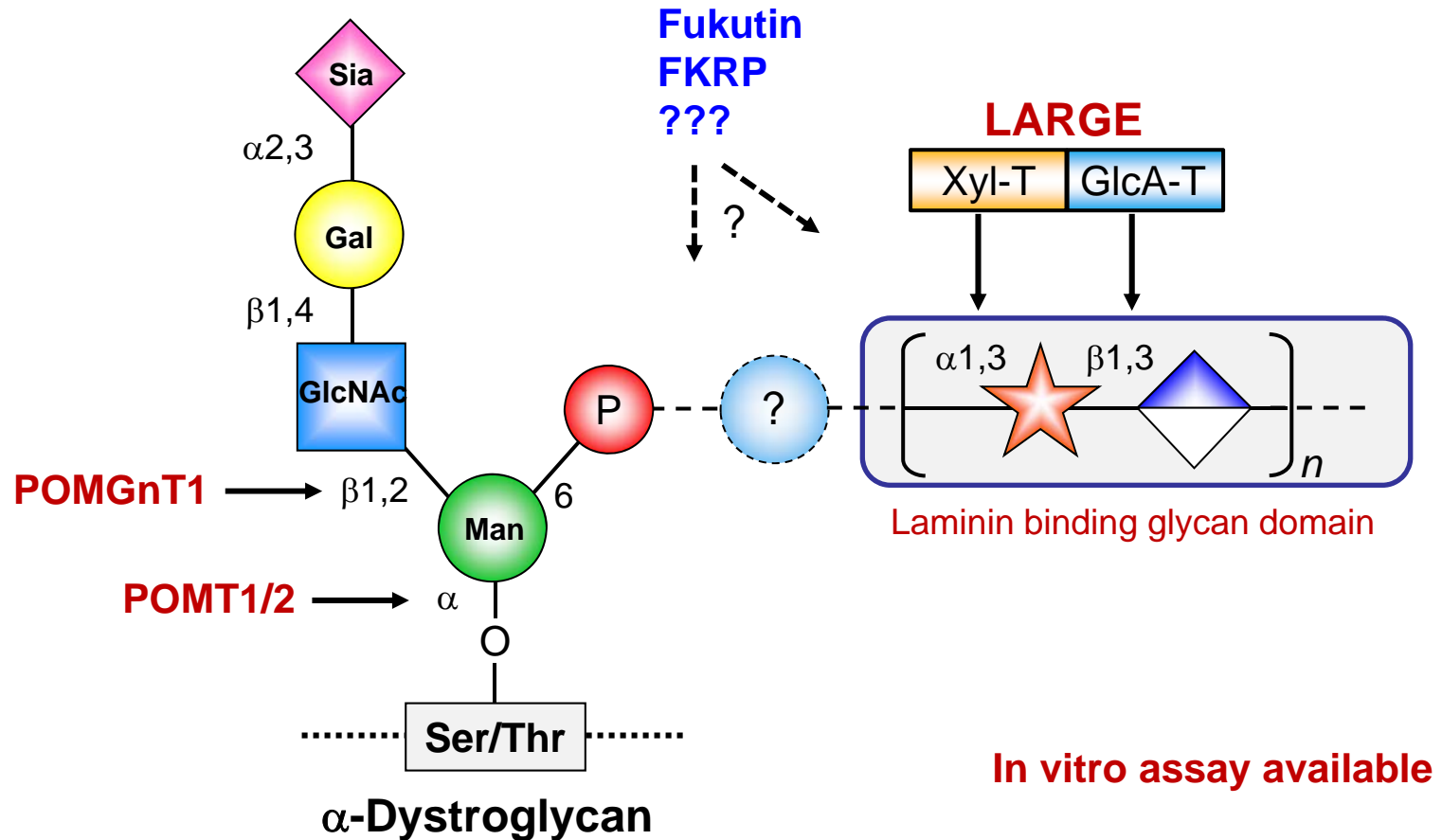
Where does ISPD fit into the α -dystroglycan glycosylation assembly line ?

- What is the function of ISPD ?
- How do *ISPD* defects affect α -DG glycosylation ?
- What step in the sugar synthesis is affected by ISPD defects ?

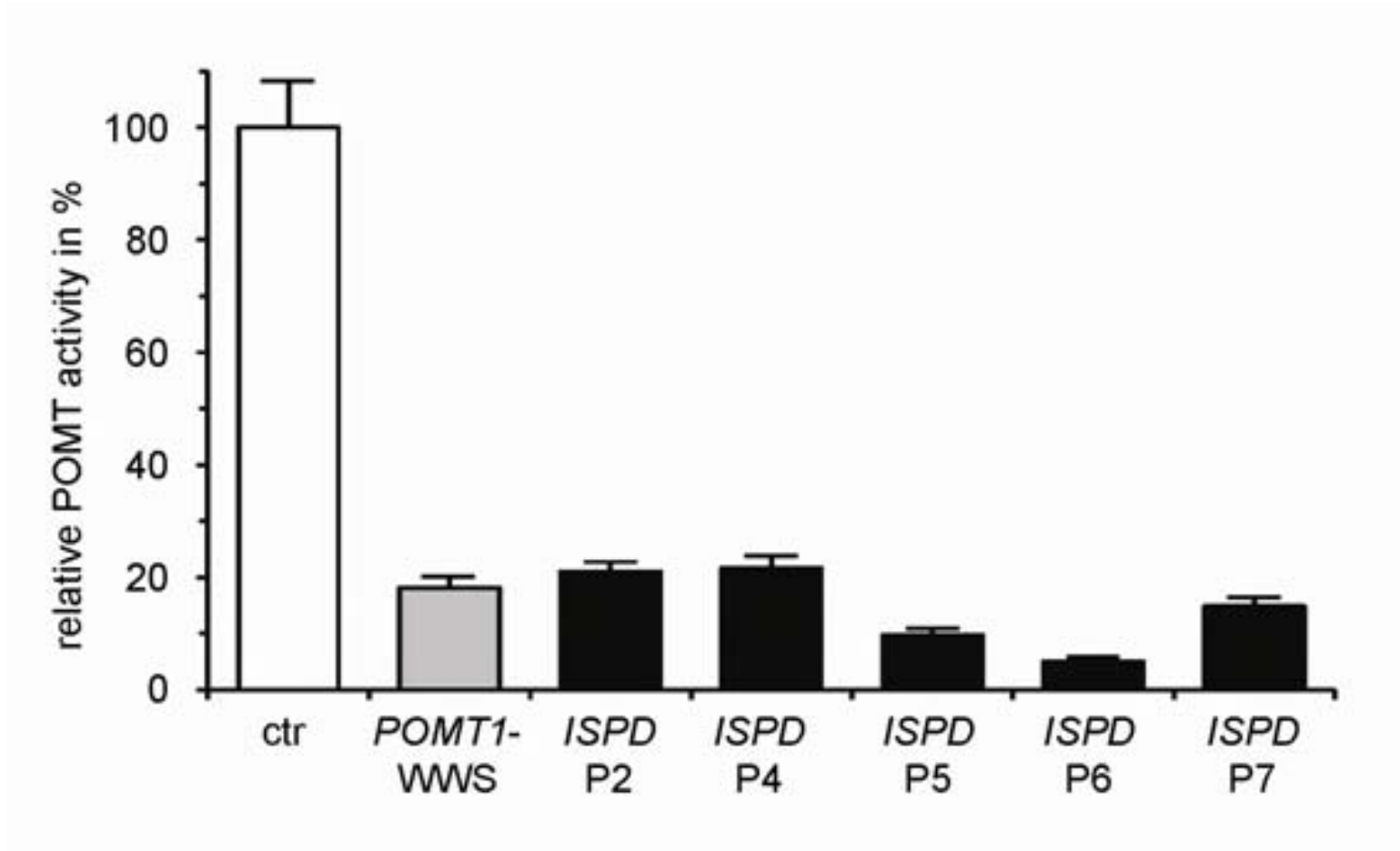


How do *ISPD* defects affect α -DG glycosylation ?

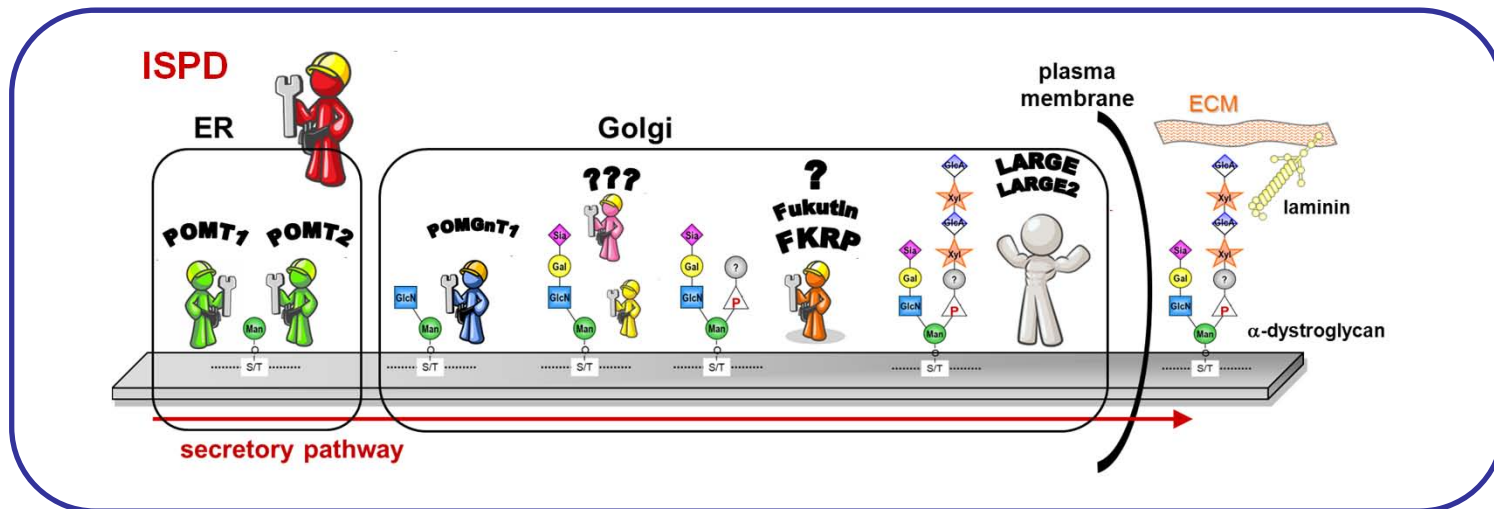
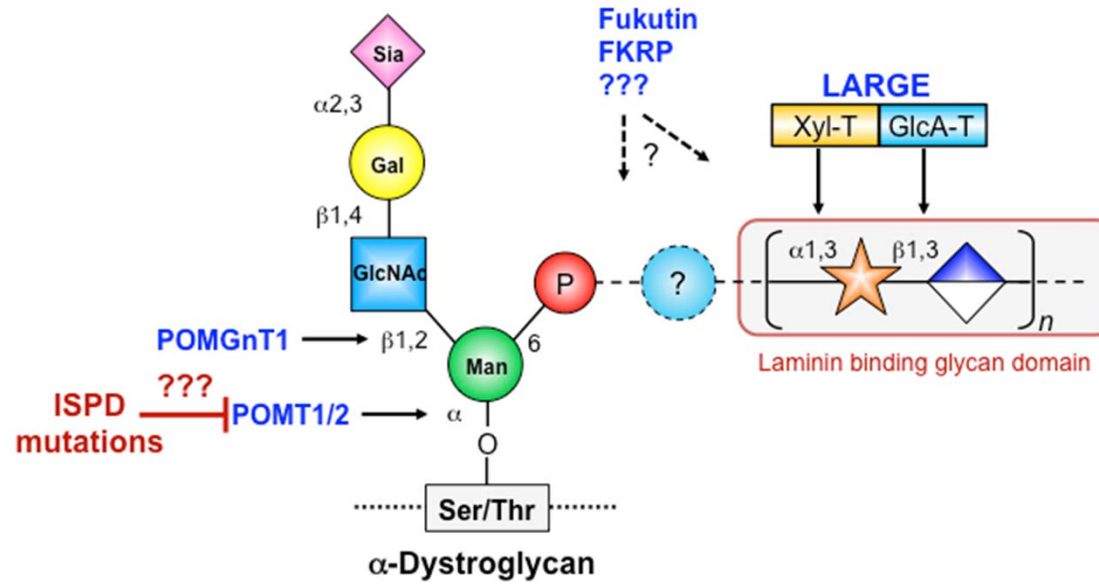
What step in the sugar synthesis is affected by *ISPD* defects ?



ISPD mutations impair protein O-mannosylation



ISPD mutations impair protein O-mannosylation



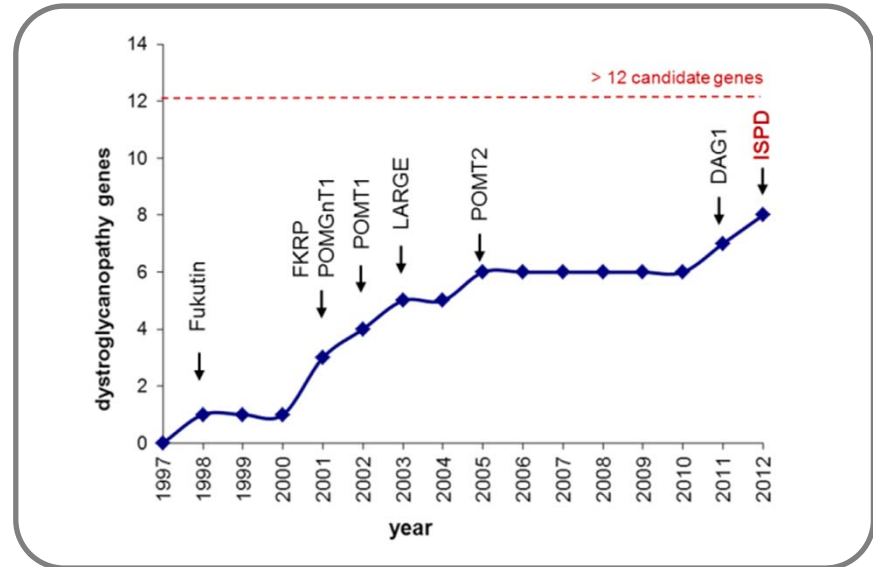
Summary

- Patient fibroblasts can be used to study α -dystroglycan glycosylation and complementation assay can be used to diagnose/validate genetic defect
- Identification of 5 novel WWS complementation groups representing 5 new WWS candidate genes
- Identification of *ISPD* gene defects as common cause in muscular dystrophies associated with α -dystroglycan glycosylation defect
- *ISPD* mutations lead to impaired α -dystroglycan O-mannosylation, establishing a new pathway and mechanism for disease in WWS.

Outlook

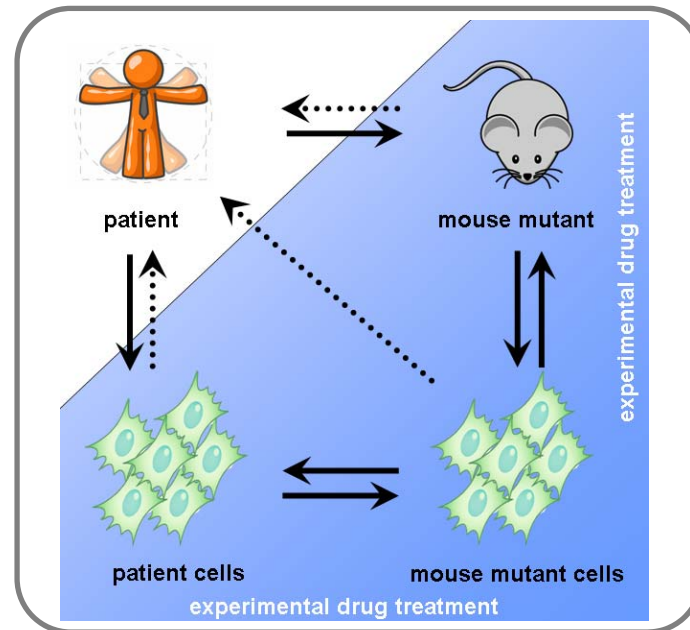
New dystroglycanopathy gene discovery:

Identify genetic defect in the remaining unidentified 4 WWS complementation groups



Screen for therapeutic compounds:

test in cell culture and mouse model systems



Acknowledgement

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Mark Lommel / Sabine Strahl

University of Heidelberg, Germany

Adeno vector generation

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